

### **Perspective**

# Innate immunity and interferon in SARS-CoV-2 infection outcome

### Ram Savan<sup>1</sup> and Michael Gale, Jr.<sup>1,\*</sup>

<sup>1</sup>Department of Immunology and Center for Innate Immunity and Immune Disease, University of Washington, 750 Republican St., Seattle, WA 98109, USA \*Correspondence: mgale@uw.edu

https://doi.org/10.1016/j.immuni.2023.06.018

### SUMMARY

Innate immunity and the actions of type I and III interferons (IFNs) are essential for protection from SARS-CoV-2 and COVID-19. Each is induced in response to infection and serves to restrict viral replication and spread while directing the polarization and modulation of the adaptive immune response. Owing to the distribution of their specific receptors, type I and III IFNs, respectively, impart systemic and local actions. Therapeutic IFN has been administered to combat COVID-19 but with differential outcomes when given early or late in infection. In this perspective, we sort out the role of innate immunity and complex actions of IFNs in the context of SARS-CoV-2 infection and COVID-19. We conclude that IFNs are a beneficial component of innate immunity that has mediated natural clearance of infection in over 700 million people. Therapeutic induction of innate immunity and use of IFN should be featured in strategies to treat acute SARS-CoV-2 infection in people at risk for severe COVID-19.

### INTRODUCTION

Nearly 700 million people worldwide have been infected with the severe acute respiratory coronavirus 2 (SARS-CoV-2), the cause of coronavirus disease 2019 (COVID-19) (https://covid19.who. int/). 6.8 million people are known to have died from infection. Vaccination for SARS-CoV-2 clearly protects against COVID-19 severity, wherein humoral immunity is the major biomarker and mediator of disease protection.<sup>1</sup> Importantly, prior to vaccination and in nonvaccinated persons today, the great majority of people infected with SARS-CoV-2 recovered following asymptomatic infection or having only acute mild to moderate disease, with recovery occurring prior to the onset or peak of humoral immunity.<sup>2,3</sup> The kinetics of infection with recovery in these COVID-19 cases, and that many cases are asymptomatic, underscore a major role for innate immunity in early control of SARS-CoV-2 infection.

### **INNATE IMMUNE CONTROL OF SARS-CoV-2**

Innate immunity provides our first line of defense against virus infection and is triggered in the infected cell through the actions of pathogen recognition receptors (PRRs) that recognize and bind specific pathogen-associated molecular patterns (PAMPs) within viral products to signal downstream innate immune activation.<sup>4</sup> In lung epithelial cells, the melanoma differentiation antigen 5 (MDA5) protein, a member of the RIG-l-like receptor (RLR) family of PRRs, recognizes SARS-CoV-2 likely through binding of viral RNA PAMPs and signals through the RLR pathway for induction of innate immunity.<sup>5–7</sup> Additional innate immune programs, possibly directed by toll-like receptors (TLRs) or other PRRs, may also contribute to innate immune activation. Innate immune activation occurs rapidly



following infection in a host cell. This process first directs the rapid expression of virus-stimulated genes (VSGs) triggered by interferon-regulatory factor 3 (IRF3), NF-KB, and other transcription factors activated by PRR signaling.<sup>8</sup> SARS-CoV-2-infected patients with rare inborn errors leading to TLR3 and TLR7 deficiencies have resulted in severe COVID-19 outcomes.<sup>9-11</sup> Further polymorphisms in transcription factors including IRF7 and IRF9 result in severe COVID-19.<sup>11</sup> These observations underscore the role of PRRs and innate immune signaling and response in early control of SARS-CoV-2. Among the VSGs are types I and III interferons (IFNs) whose production and secretion lead to IFN signaling in the infected cell and neighboring cells to induce a major second wave of gene expression by directing the induction of hundreds of interferon-stimulated genes (ISGs) and a tissue-wide and even systemic antiviral state (Figure 1). While VSG products provide an initial wave of gene expression for first-line defense to limit local virus replication and spread, ISGs expand and diversify this response to systemically mediate antiviral defenses.<sup>12</sup> VSG and ISG products also include specific chemokines and cytokines that serve to program and polarize the adaptive immune response for effective immunity and viral clearance. Cytokine and chemokine products such as interleukin-6 (IL-6), tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ), and CXCL10 are also implicated in the pathogenic cytokine storm underlying severe COVID-19 outcomes,<sup>13,14</sup> so the innate immune response must be fine-tuned to direct recovery versus disease.<sup>15,16</sup> Moreover, viral proteins that accumulate in the infected cell during the course of SARS-CoV-2 replication can attenuate innate immunity (described below and reviewed elsewhere<sup>17</sup>), thus marking the rapid onset and actions of innate immunity as key features in controlling infection and protecting against development and severity of COVID-19.





# CONTRASTING ROLES OF TYPES I AND III IFNs IN SARS-CoV-2 CLEARANCE

While types I and III IFNs are induced through the same PRRs, their transcriptional response kinetics are distinct and should be expected to impart temporally distinct antiviral actions against SARS-CoV-2. Type I IFNs induce ISG expression early following receptor signaling, while type III IFN-induced ISGs are comparatively delayed, but their expression is sustained for a longer period.<sup>18,19</sup> While both IFNs activate the Jak-STAT signaling cascade to induce ISGs (see Figure 1), type I IFNs alone can also activate STAT1 homodimers to induce inflammatory signals, thus driving a different composition of ISGs.<sup>18</sup> Further, type III IFN receptor expression is restricted to epithelial layers at the mucosal surfaces compared to the ubiquitous expression of type I IFN receptor. These differences impart a division of labor, with type I IFNs playing a dominant role in inducing broad tissue and systemic responses and type III IFNs activating innate antiviral responses at the mucosal tissues.

Primary infection of nasal and lung epithelium marks SARS-CoV-2 as a mucosal virus.<sup>20</sup> As type III IFN responses are mainly confined to surfaces of the mucosal epithelium, this IFN is critical to control infection by mucosal viruses including norovirus,<sup>21–23</sup> respiratory syncytial virus, influenza A virus (IAV),<sup>24</sup> and SARS-CoV-2.<sup>25</sup> The SARS-CoV-2/COVID-19 pandemic has drawn new attention to the ongoing debate on IFNs as therapeutic agents, particularly the role of type III IFN in the lung. Severe COVID-19 patients have high levels of circulating IFNs, thereby associating IFNs with a worse prognosis of the disease<sup>26</sup> (**Figure 2**). To address the mechanisms of bad outcomes due to high, chronic levels of IFNs *in vivo*, studies have evaluated mucosal integrity in mouse models in the context of high-dose IAV or treatment with poly I:C (a double-stranded RNA PAMP



# Figure 1. SARS-CoV-2 induction of innate immune activation and IFNs

SAR-CoV-2 infection and cell entry results in viral RNA deposition in the cell cytosol to initiate viral protein synthesis and viral RNA replication. Left: MDA5 first senses viral products. likely viral RNA components, and undergoes signaling activation involving interaction with the MAVS adaptor protein. SARS-CoV-2 might also be sensed through other pathogen recognition receptors including specific Toll-like receptors. Signaling mediates downstream activation and nuclear accumulation of IRF3, NF-KB, and other transcription factors that direct the expression of virus-stimulated genes (VSGs), many with antiviral and immunemodulatory activity. Right: types I and III IFNs are cytokine VSG products and are secreted from the infected cell and signal via distinct receptors engaging the Jak-STAT pathway, leading to interferon-stimulated gene (ISG) expression. Hundreds of ISGs are induced in lung epithelial cells. ISG actions amplify and diversify the innate immune response, with the responses to type I and type III IFNs being temporally distinct and respectively partitioned by systemic versus compartmentalized (type III IFN) receptor expression distribution. Specific ISGs mediate antiviral actions against SARS-CoV-2. IFN therapy strategies leverage these actions and the immunomodulatory activities of ISGs for the treatment of infection and COVID-19.

commonly used to induce IFN expression) to model SARS-CoV-2 infection.<sup>27-29</sup> The induced and chronic IFNs delayed epithelial repair of damage caused by virus infection or PAMPinduced inflammatory response in the mucosa. By contrast, pre-infection or early treatment of mice with type III IFN was protective against disease following virus challenge. Thus, location, timing, and duration of IFN exposure determines the protective or detrimental effects of IFN actions. More recent studies have clarified the role of type III IFN using mouse-adapted strains of SARS-CoV-2 in the mouse model of infection. Here, type III IFN protects mice challenged with SARS-CoV-2 Alpha and Omicron variants, protecting against virus-induced inflammation and tissue damage in both the upper and lower airways.<sup>25</sup> Early type III IFN intranasal administration to mice can also provide protection from morbidity and mortality otherwise caused by SARS-CoV-2, indicating that the timing of IFN production and signaling are critical for protection against disease.<sup>25</sup> These outcomes align with previous studies from us and others showing that type III IFN induces tissue-specific immunity without causing damaging inflammation, as type III IFN does not recruit immune cells to the site of infection.<sup>18,30,31</sup>

There are several contrasting studies regarding the role of type I IFNs in the outcome of experimental SARS-CoV-2 infection. Considering the analyses of SARS-CoV-2 mouse models of infection utilizing the transgenic expression of human angiotensin-converting enzyme 2 (ACE2) in mice, some studies show that the response to IFN links with control of infection in this model,<sup>25,32</sup> while others document enhanced lung inflammation caused by type I IFN linked with lack of virus clearance.<sup>33</sup> These contrasting outcomes can be due to differences in virus challenge dose and route of challenge and should consider the impact of tissue distribution of transgenic human ACE2 expression, which is aberrantly expressed in a range of tissues in this model.<sup>34</sup> However, we note that previous studies of the

### Immunity

Perspective



MERS-CoV have shown that IFNs can mediate protective actions *in vivo*.<sup>35</sup> These observations warrant an in-depth biological characterization of IFN actions against SARS-CoV-2, taking routes of infection, dose, cell type, and location-specific depletion of IFN receptors into consideration. While more studies are needed to characterize the role of type I IFNs in the context of SARS-CoV-2, it is clear from other virus infections that they are critical to early virus control and initiating/modulating adaptive immune responses.<sup>36</sup> We propose that types I and III IFNs play a collaborative but non-redundant role in protecting against SARS-CoV-2 and COVID-19.

### SPECIFIC ISGs RESTRICTING SARS-CoV-2

IFNs mediate their antiviral actions through the function of ISGs, including genes that are differentially expressed by types I and III IFNs.<sup>37</sup> Most ISGs described in the literature are those whose expression is induced in response to IFNs, but it should be noted that there are many ISGs whose expression is reduced or down-regulated from pre-IFN baseline levels.<sup>12,38</sup> While induced ISGs can impact viral replication directly, the reduced expression ISGs include genes encoding factors that regulate metabolic and cell growth control to confer restriction of virus infection,<sup>38</sup> but the spectrum of the ISGs impacting SARS-CoV-2 is not defined. The antiviral response linked with ISGs is designed to affect every step of the virus life cycle, with specific ISGs thus far shown to restrict SARS-CoV-2 entry or viral RNA replication, but restriction factors affecting SARS-CoV-2 egress have



## Figure 2. IFN actions against SARS-CoV-2 mediated by select ISGs

Cell infection progression of SARS-CoV-2 is depicted. In the context of IFN production or treatment, types I and III IFNs signal through the Jak-STAT pathway and induce the assembly, nuclear accumulation, and activity of IFNstimulated gene factor 3 (ISGF3) transcription factor, which binds to the interferon-stimulated response element (ISRE) promoter region of ISGs, including OAS1, ISG15, MX1, are induced by IFNs. Major ISGs that suppress/restrict SARS-CoV-2 replication at specific steps in the viral life cycle are shown.

not been identified (Figure 2). Some specific ISGs restricting SARS-CoV-2 are featured below.

SARS-CoV-2 can enter the cell through two main mechanisms, ACE2-mediated endosomal entry or TMPRSS2-mediated cell-surface entry via membrane fusion. The major mode of SARS-CoV-2 entry is through TMPRSS2-mediated activation/ cleavage of Spike to an S2 cleavage product facilitating binding to Ace2.<sup>39</sup> Among ISG products, LY6E was found to restrict S-protein-mediated membrane entry.<sup>40,41</sup> The effect of LY6E diverges beyond SARS-CoV-2, as mice lacking LY6E were protected against another co-

ronavirus—the mouse hepatitis virus (MHV). This observation is intriguing, given that LY6E has proviral functions in the context of flaviviruses, IAV, and HIV.<sup>42–44</sup> Dissecting the underlying molecular mechanisms of how LY6E mediates its activity to restrict coronaviruses will shed light on these seemingly opposing virus-specific functions.

A transposon screen to identify restriction factors of the Ebola virus identified a specific isoform of CD74 that importantly also inhibits the cleavage of the SARS-CoV-2 S protein, disrupting its processing in the endosome.<sup>45</sup> The p41 isoform of CD74 inhibited viral entry by blocking cathepsin-mediated processing of the Ebola virus glycoprotein. Remarkably, CD74 p41 can also block the endosomal entry pathway of coronaviruses, including SARS-CoV-2, through actions induced by CIITA, an ISG and transcription co-factor of major histocompatibility (MHC) expression and upstream regulator of CD74.45 Thus, CIITA and CD74 impart an IFN-induced pathway of ISG actions against SARS-CoV-2 entry. Moreover, multiple studies have implicated IFN-induced transmembrane protein 3 (IFITM3) in SARS-CoV-2 entry restriction.<sup>46,47</sup> IFITM3, a well-studied restriction factor against IAV, HIV, and West Nile virus, inhibits the fusion of viral and cellular membranes to preclude virus entry into target cells.<sup>48</sup> In the case of SARS-CoV-2, the IFITM3 protein binds to the virus, blocking viral entry. A single-nucleotide polymorphism (SNP) of IFITM3 is linked to increased susceptibility to SARS-CoV-2,<sup>49</sup> with the potential of utilizing this SNP to identify individuals who are more likely to be susceptible to the virus.



OAS1 is one of the major SARS-CoV-2 restriction factors that could partially explain the heterogeneity in innate mediated control of SARS-CoV-2 infection in susceptible people. A 46-kDa splice isoform of oligoadenylate synthetase 1 (OAS1), called OAS1p46 or prenylated OAS1, displayed robust antiviral activity against SARS-CoV-2. OAS proteins are IFN-inducible innate immune proteins that sense and bind to viral RNA and activate a ribonuclease RNaseL to control virus replication.50-53 The antiviral activity of OAS1p46 against SARS-CoV-2 is dependent on prenylation of OAS1p46. Prenylated OAS1 traffics to the Golgi, including possibly the SARS-CoV-2 double-membrane replication organelle (Figure 2).54,55 Interestingly, the human OAS1 gene produces two major isoforms, OAS1p42 or OAS1p46 (prenylated isoform), and their expression is controlled by an SNP in the sixth exon splice site acceptor (A/G; rs10774671) of the OAS1 gene. The G allele generates the OAS1p46 isoform, and the A allele generates OAS1p42. Both OAS1p42 and OAS1p46 are expressed if an individual carries A and G alleles. Individuals who carry homozygous alleles naturally lack either OAS1p42 (G/G) or OAS1p46 (A/A) isoforms, while heterozygotes express both isoforms. In the context of COVID-19, A/A homozygous patients who are OAS1p46 deficient progress to severe COVID-19, requiring mechanical ventilation or leading to death.55 Recent studies have replicated these virological and genetic findings, implicating prenylated OAS1 as a central restriction factor of SARS-CoV-2, whose loss of function links with progression to severe COVID-19.56,57 While genetic variation in OAS1 has emerged as a significant disease predictor in multiple COVID-19 patient cohorts, it is still puzzling that the odds ratio linked to this SNP is not higher, which suggests that there are other variables affecting the function of prenylated OAS1 against SARS-CoV-2. As early innate immune-mediated control of SARS-CoV-2 is critical in preventing the infected individual from progressing to severe COVID-19, we presume that prenylated OAS1 expression is directly affected by the level of IFNs induced and hence the actions of other ISGs during actute SARS-CoV-2 infection. We propose that the SNPs in genes operating in the IFN signaling pathway<sup>58</sup> or IFN autoantibodies<sup>59-61</sup> (see below) could differentially influence the protective effects of prenylated OAS1. These observations underscore the significance of prenylated OAS1 in restricting SARS-CoV-2.

Zinc finger antiviral protein (ZAP) is another ISG that restricts SARS-CoV-2. The Zap gene produces multiple splice isoforms, with predominant expression of long (ZAP-L) and short (ZAP-S) isoforms.<sup>54</sup> ZAP-L is constitutively expressed, while ZAP-S is an ISG. SARS-CoV-2 studies have implicated both isoforms as playing a role in virus restriction, with each mediating distinct mechanisms of action against the virus. Additionally, another study shows that ZAP-L mediates virus restriction through RNase activity, requiring KHYHN and TRIM25 as accessory factors, 62 with ZAP potently restricting SARS-CoV-2. This work also shows that ZAP-L enforces constitutive antiviral activity, with ZAP-S directing IFN-induced antiviral actions. Another study implicates ZAP-S in restricting SARS-CoV-2 by interfering with ribosomal frameshifting, thereby inhibiting viral replication.<sup>63</sup> Thus, ZAP isoforms mediate basal and IFNinduced antiviral actions against SARS-CoV-2 (Figure 2).

While the afore described ISGs are shown to have defined functions to mediate IFN antiviral actions against SARS-CoV-2,

### Immunity Perspective

the spectrum of antiviral ISGs that restrict SARS-CoV2 is yet to be identified. CRISPR and mass spectrometry screens have revealed several host factors that directly or indirectly interact with SARS-CoV-2 products or whose expression levels impact viral production.<sup>64–73</sup> Defined host factors from these studies are shown to impact SARS-CoV-2 processes of viral entry, replication, and virion morphogenesis and to regulate organelle modifications during infection, thus impacting the cellular homeostasis. We note that several of these factors could also have pro-viral functions, direct or indirect linkage with ISGs and IFN actions, and could represent novel antiviral programs of innate immunity. Important next steps from these multiple studies are to validate the "hits" from each screen to identify the gene, protein, and signaling networks that confer mechanism of action to regulate SARS-CoV-2 infection outcome.

# PERTURBATIONS OF INTERFERON SIGNALING AND IMPLICATIONS TO SARS-CoV-2 CONTROL

Type I IFNs signal through IFN $\alpha/\beta$  receptors (IFNAR), which are expressed on nucleated cells. Humans express 14 subtypes of type I IFNs with varying affinity to the IFN receptors.<sup>36</sup> Only IFN $\alpha$  and IFN $\beta$  are produced in the lungs during SARS-CoV-2 infection. Although type I IFNs induce an early but transient innate antiviral program, they are also critical for promoting the adaptive immune response. For example, IFNs activate antigen-presenting cells and cooperate with other cytokines to polarize T cells, mediate induction of the immuno-proteasome, and enhance expression of mMHC molecules on the surface of cells to facilitate antigen presentation and T cell-mediated killing of infected cells, respectively.<sup>74</sup> In virus infection, the absence of IFN is catastrophic for the host cell and organism.<sup>36</sup> As such, IFNAR-deficient mice exhibit increased susceptibility and tissue pathology compared to wild-type mice when infected with mouse-adapted SARS-CoV-2.75 Moreover, recent studies uncovered autoantibodies against type I IFNs in people. with an implication that the autoantibodies will block IFNs from binding the IFN receptor and activate downstream signaling<sup>76,77</sup> (see Figure 1). These important studies show that 10% of severe COVID-19 patients carry IFN autoantibodies that attenuate IFN signaling and ISG expression.<sup>78</sup> Notably, the prevalence of IFN autoantibodies is higher in older individuals, and SARS-CoV-2 patients harboring such autoantibodies progress to severe COVID-19 or mortality.77 These findings reveal an important role for IFN actions in recovery from SARS-CoV-2 infection and mitigation of COVID-19 and are broadly relevant to other virus infections. Importantly, the presence of type III IFN autoantibodies in patients is not as prevalent as for type I IFN autoantibodies. If this differential observation holds up in multiple cohorts, this finding could have significant biological implications, including the use of recombinant type III IFNs as therapy against SARS-CoV-2 infections. Human genetic variations also have a significant biological effect on IFN signaling and COVID-19 severity. Recent genome-wide association studies revealed polymorphisms in IFNAR2 and TYK2 genes that affect COVID-19 severity.58 While further studies are needed to decipher the mechanisms of action underlying disease susceptibility in this genetic context, human variations in the IFN signaling pathway explain some of the heterogeneity

### Immunity Perspective

in the outcome of acute SARS-CoV-2 infection and clearly implicate IFN as crucial for innate immune defense and protection from COVID-19.

### **IFN EVASION BY SARS-CoV-2**

Like other pathogenic viruses, SARS-CoV-2 encodes proteins that can suppress IFN induction and actions This topic has been reviewed extensively (see Minkoff and tenOever<sup>17</sup> for example) but without context of the temporal nature of innate immune activation and the SARS-CoV-2 replication cycle. We should make it clear that for any virus, initial infection launches an arms race for the host to induce innate immune defenses and IFNs to suppress virus replication and spread, wherein the virus must strike with countermeasures to evade and/or actively suppress innate immune and IFN actions of the host cell in order to survive, make more virions, and spread to new hosts.<sup>36</sup> Disease is a side effect of these collective host and virus actions. SARS-CoV-2 encodes proteins reported to suppress PRR signaling, block protein nuclear import that impacts the actions of activated IRF3 and STAT proteins, and suppress specific ISG functions.<sup>17</sup> Many studies that have described innate immune and IFN evasion actions of SARS-CoV-2 to date have relied on cell culture models of viral protein overexpression conducted without context of actual SARS-CoV-2 infection and/or without consideration of host and virus response kinetics, including the kinetics of VSG and ISG expression and function. Studies to model infection in relevant cells of the upper and lower airway mucosa and across specific tissues of infection spread are required. Additionally, studies to document effects of wild-type and mutant SARS-CoV-2 engineered with specific protein-expression/functional deletions, as well as across contemporary viral variants, are needed. Such studies are essential to ascertain the true actions of viral proteins in evasion from innate immune and IFN defenses given that these innate defenses facilitate viral control and recovery from infection in most people exposed to SARS-CoV-2, even in the era of widespread vaccination. It is also important to note that as new SARS-CoV-2 variants continue to emerge and escape vaccine immunity,79 innate immune and IFN defenses remain our foundational immune defense against new virus exposure and infection, linking with hybrid immunity to suppress infection and progression to COVID-19.80,81 Continued research to define the virus-host interactions of innate immune and IFN control of infection is needed to ascertain the spectrum of host genes that serve to innately protect us against SARS-CoV-2 infection and disease and to understand how these processes regulate the contemporary status of hybrid immunity to infection. Such efforts should reveal the spectrum VSG and ISG targets and their upstream regulatory processes for consideration in strategies of host-directed therapeutics to suppress SARS-CoV-2 and enhance hybrid immunity.

#### **IFN TREATMENT AND OUTCOMES**

The clinical use of IFNs against SARS-CoV-2 has been met with skepticism because of the history of varied outcomes while treating cancer, autoimmunity, and other virus infections. A major barrier to the clinical use of type I IFNs are the side effects that include flu-like symptoms, anemia, and depression.<sup>82,83</sup> Multiple



randomized controlled clinical trials have now evaluated IFN therapy for improving outcomes in SARS-CoV-2 infection (summarized in Ryoo et al.<sup>84</sup> and Chen et al.<sup>85</sup>). These include studies of systemic or aerosol/inhaled administration of IFN, which it is important to note was typically administered late in infection to hospitalized patients with severe COVID-19. In clinical trials of IFNα treatment, there was no significant difference in mortality from placebo control across COVID-19 cohorts, but treatment was indeed linked with a significant increase in the number of patients who improved enough to be discharged from the hospital.<sup>86</sup> Moreover, the administration of IFN $\beta$  did not increase the survival of severe COVID-19 cases among hospitalized patients, but it did impact the disease to reduce the need for intensive care.<sup>85</sup> In a departure from treating hospitalized patients with IFN, two clinical trials of an early, systemic single administration of pegylated (long-lived) type III IFN evaluated efficacy for reducing acute phase virus and suppressing disease onset among outpatients. In one study, the cohorts included outpatients with laboratory-confirmed SARS-CoV-2 within 7 days of symptom onset and asymptomatic outpatients testing positive or within 7 days of study.<sup>87</sup> Type III IFN administration was found to increase the number of outpatients with viral clearance by day 7, including individuals with the highest initial viral load. Thus, early administration of type III IFN is beneficial against SARS-CoV-2. Similar observations were recorded in another study where pegylated IFNs were given to mostly vaccinated although infected with SARS-CoV-2 outpatients.<sup>88</sup> All IFNs were shown to be safe and tolerable across these studies, with some adverse events reported. Taken together, these clinical studies show that late administration of IFN does not impact survival in cases of severe COVID-19 but does significantly link with clinical case improvement that can reduce hospital stay or entry into intensive care units. Most importantly, early administration of IFN (shown by type III IFN) during acute infection clearly presents the opportunity to reduce viral load and hence virus shedding and transmission of SARS-CoV-2, outcomes that mitigate disease progression and interpersonal spreading of the virus.

#### CONCLUSIONS

IFNs are critical components of innate immune defense and immune programming against virus infection. The actions of IFNs are mediated through hundreds of ISGs, the function of most of which are not defined. A growing list of VSGs and ISGs are now known to mediate antiviral actions that suppress specific stages of SARS-CoV-2 infection/replication but continued work in this area is sorely needed. IFNs are among the VSGs that are induced during acute virus infection and are indeed produced in response to SARS-CoV-2 infection despite viral counter measures to suppress IFN production from infected cells. Most people infected with SARS-CoV-2 have little to moderate disease and recover from infection owing to the antiviral actions the innate immune response and IFNs. While IFN levels in the blood are linked with inflammatory disease and worse COVID-19 in hospitalized patients, one cannot attribute COVID-19 severity to IFNs among the complex inflammatory response underlying COVID-19. In our opinion, IFN remains a viable therapeutic for the early treatment of acute SARS-CoV-2 infection. Placement of aerosol or systemic IFN treatments into clinical strategies aimed at



protecting people at high risk for severe COVID-19, with IFN treatment administration occurring as soon as possible after testing virus positive, would induce an antiviral response to reduce the duration of viral shedding, suppress viral load, and suppress or mitigate disease. Finally, research to define innate immune and IFN actions in the growing context of global/continued vaccination, emergence of vaccine-escape viral variants, and hybrid immunity is needed to define therapeutic strategies for control of SARS-CoV-2 infection and to inform preparation for future epidemics and pandemics from emerging viruses.

#### ACKNOWLEDGMENTS

We were unable to cite all of the great many studies of innate immunity and SARS-CoV-2, and we thank our colleagues for their terrific work in this area. This work was supported by National Institutes of health grants Al151698 and Al148684 to M.G. and Al175724 and Al176442 to R.S.

#### **DECLARATION OF INTERESTS**

The authors declare no competing interests.

#### REFERENCES

- Agyapon-Ntra, K., and McSharry, P.E. (2023). A global analysis of the effectiveness of policy responses to COVID-19. Sci. Rep. 13, 5629. https://doi.org/10.1038/s41598-023-31709-2.
- Drain, P.K., Dalmat, R.R., Hao, L., Bemer, M.J., Budiawan, E., Morton, J.F., Ireton, R.C., Hsiang, T.Y., Marfatia, Z., Prabhu, R., et al. (2023). Duration of viral infectiousness and correlation with symptoms and diagnostic testing in non-hospitalized adults during acute SARS-CoV-2 infection: A longitudinal cohort study. J. Clin. Virol. *161*, 105420. https://doi.org/10. 1016/j.jcv.2023.105420.
- Wei, X., Narasimhan, H., Zhu, B., and Sun, J. (2023). Host Recovery from Respiratory Viral Infection. Annu. Rev. Immunol. 41, 277–300. https://doi. org/10.1146/annurev-immunol-101921-040450.
- Chow, K.T., Gale, M., Jr., and Loo, Y.M. (2018). RIG-I and Other RNA Sensors in Antiviral Immunity. Annu. Rev. Immunol. 36, 667–694. https://doi.org/10.1146/annurev-immunol-042617-053309.
- Rebendenne, A., Valadão, A.L.C., Tauziet, M., Maarifi, G., Bonaventure, B., McKellar, J., Planès, R., Nisole, S., Arnaud-Arnould, M., Moncorgé, O., and Goujon, C. (2021). SARS-CoV-2 triggers an MDA-5-dependent interferon response which is unable to control replication in lung epithelial cells. J. Virol. 95, e02415-20. https://doi.org/10.1128/JVI.02415-20.
- Yin, X., Riva, L., Pu, Y., Martin-Sancho, L., Kanamune, J., Yamamoto, Y., Sakai, K., Gotoh, S., Miorin, L., De Jesus, P.D., et al. (2021). MDA5 Governs the Innate Immune Response to SARS-CoV-2 in Lung Epithelial Cells. Cell Rep. 34, 108628. https://doi.org/10.1016/j.celrep.2020.108628.
- Rehwinkel, J., and Gack, M.U. (2020). RIG-I-like receptors: their regulation and roles in RNA sensing. Nat. Rev. Immunol. 20, 537–551. https://doi.org/ 10.1038/s41577-020-0288-3.
- Andersen, J., VanScoy, S., Cheng, T.F., Gomez, D., and Reich, N.C. (2008). IRF-3-dependent and augmented target genes during viral infection. Genes Immun. 9, 168–175. https://doi.org/10.1038/sj.gene.6364449.
- Asano, T., Boisson, B., Onodi, F., Matuozzo, D., Moncada-Velez, M., Maglorius Renkilaraj, M.R.L., Zhang, P., Meertens, L., Bolze, A., Materna, M., et al. (2021). X-linked recessive TLR7 deficiency in ~1% of men under 60 years old with life-threatening COVID-19. Sci. Immunol. 6, eabl4348. https://doi.org/10.1126/sciimmunol.abl4348.
- Fallerini, C., Daga, S., Mantovani, S., Benetti, E., Picchiotti, N., Francisci, D., Paciosi, F., Schiaroli, E., Baldassarri, M., Fava, F., et al. (2021). Association of Toll-like receptor 7 variants with life-threatening COVID-19 disease in males: findings from a nested case-control study. Elife 10, e67569. https://doi.org/10.7554/eLife.67569.



- Zhang, Q., Bastard, P., Liu, Z., Le Pen, J., Moncada-Velez, M., Chen, J., Ogishi, M., Sabli, I.K.D., Hodeib, S., Korol, C., et al. (2020). Inborn errors of type I IFN immunity in patients with life-threatening COVID-19. Science 370, eabd4570. https://doi.org/10.1126/science.abd4570.
- Green, R., Ireton, R.C., and Gale, M., Jr. (2018). Interferon-stimulated genes: new platforms and computational approaches. Mamm. Genome 29, 593–602. https://doi.org/10.1007/s00335-018-9755-6.
- Lorè, N.I., De Lorenzo, R., Rancoita, P.M.V., Cugnata, F., Agresti, A., Benedetti, F., Bianchi, M.E., Bonini, C., Capobianco, A., Conte, C., et al. (2021). CXCL10 levels at hospital admission predict COVID-19 outcome: hierarchical assessment of 53 putative inflammatory biomarkers in an observational study. Mol. Med. 27, 129. https://doi.org/10.1186/s10020-021-00390-4.
- Del Valle, D.M., Kim-Schulze, S., Huang, H.H., Beckmann, N.D., Nirenberg, S., Wang, B., Lavin, Y., Swartz, T.H., Madduri, D., Stock, A., et al. (2020). An inflammatory cytokine signature predicts COVID-19 severity and survival. Nat. Med. 26, 1636–1643. https://doi.org/10.1038/s41591-020-1051-9.
- Ahmed, D., Al-Daraawi, M., and Cassol, E. (2023). Innate sensing and cellular metabolism: role in fine tuning antiviral immune responses. J. Leukoc. Biol. 113, 164–190. https://doi.org/10.1093/jleuko/giac011.
- Anaeigoudari, A., Mollaei, H.R., Arababadi, M.K., and Nosratabadi, R. (2021). Severe Acute Respiratory Syndrome Coronavirus 2: The Role of the Main Components of the Innate Immune System. Inflammation 44, 2151–2169. https://doi.org/10.1007/s10753-021-01519-7.
- Minkoff, J.M., and tenOever, B. (2023). Innate immune evasion strategies of SARS-CoV-2. Nat. Rev. Microbiol. 21, 178–194. https://doi.org/10. 1038/s41579-022-00839-1.
- Forero, A., Ozarkar, S., Li, H., Lee, C.H., Hemann, E.A., Nadjsombati, M.S., Hendricks, M.R., So, L., Green, R., Roy, C.N., et al. (2019). Differential Activation of the Transcription Factor IRF1 Underlies the Distinct Immune Responses Elicited by Type I and Type III Interferons. Immunity *51*, 451– 464.e6. https://doi.org/10.1016/j.immuni.2019.07.007.
- Dowling, J.W., and Forero, A. (2022). Beyond Good and Evil: Molecular Mechanisms of Type I and III IFN Functions. J. Immunol. 208, 247–256. https://doi.org/10.4049/jimmunol.2100707.
- Russell, M.W., and Mestecky, J. (2022). Mucosal immunity: The missing link in comprehending SARS-CoV-2 infection and transmission. Front. Immunol. 13, 957107. https://doi.org/10.3389/fimmu.2022.957107.
- Baldridge, M.T., Lee, S., Brown, J.J., McAllister, N., Urbanek, K., Dermody, T.S., Nice, T.J., and Virgin, H.W. (2017). Expression of IfnIr1 on Intestinal Epithelial Cells Is Critical to the Antiviral Effects of Interferon Lambda against Norovirus and Reovirus. J. Virol. 91, e02079-16. https:// doi.org/10.1128/JVI.02079-16.
- Baldridge, M.T., Nice, T.J., McCune, B.T., Yokoyama, C.C., Kambal, A., Wheadon, M., Diamond, M.S., Ivanova, Y., Artyomov, M., and Virgin, H.W. (2015). Commensal microbes and interferon-lambda determine persistence of enteric murine norovirus infection. Science 347, 266–269. https://doi.org/10.1126/science.1258025.
- Nice, T.J., Baldridge, M.T., McCune, B.T., Norman, J.M., Lazear, H.M., Artyomov, M., Diamond, M.S., and Virgin, H.W. (2015). Interferon-lambda cures persistent murine norovirus infection in the absence of adaptive immunity. Science 347, 269–273. https://doi.org/10.1126/science.1258100.
- Hemann, E.A., Green, R., Turnbull, J.B., Langlois, R.A., Savan, R., and Gale, M., Jr. (2019). Interferon-lambda modulates dendritic cells to facilitate T cell immunity during infection with influenza A virus. Nat. Immunol. 20, 1035–1045. https://doi.org/10.1038/s41590-019-0408-z.
- Chong, Z., Karl, C.E., Halfmann, P.J., Kawaoka, Y., Winkler, E.S., Keeler, S.P., Holtzman, M.J., Yu, J., and Diamond, M.S. (2022). Nasally delivered interferon-lambda protects mice against infection by SARS-CoV-2 variants including Omicron. Cell Rep. 39, 110799. https://doi.org/10.1016/j. celrep.2022.110799.
- da Silva, R.P., Gonçalves, J.I.B., Zanin, R.F., Schuch, F.B., and de Souza, A.P.D. (2021). Circulating Type I Interferon Levels and COVID-19 Severity: A Systematic Review and Meta-Analysis. Front. Immunol. *12*, 657363. https://doi.org/10.3389/fimmu.2021.657363.

### Immunity Perspective



- Hendricks, M.R., and Savan, R. (2020). Interferon-lambda at the Center of the Storm. Immunity 53, 245–247. https://doi.org/10.1016/j.immuni.2020. 07.024.
- Major, J., Crotta, S., Llorian, M., McCabe, T.M., Gad, H.H., Priestnall, S.L., Hartmann, R., and Wack, A. (2020). Type I and III interferons disrupt lung epithelial repair during recovery from viral infection. Science 369, 712–717. https://doi.org/10.1126/science.abc2061.
- Broggi, A., Ghosh, S., Sposito, B., Spreafico, R., Balzarini, F., Lo Cascio, A., Clementi, N., De Santis, M., Mancini, N., Granucci, F., and Zanoni, I. (2020). Type III interferons disrupt the lung epithelial barrier upon viral recognition. Science 369, 706–712. https://doi.org/10.1126/science. abc3545.
- Henden, A.S., Koyama, M., Robb, R.J., Forero, A., Kuns, R.D., Chang, K., Ensbey, K.S., Varelias, A., Kazakoff, S.H., Waddell, N., et al. (2021). IFNlambda therapy prevents severe gastrointestinal graft-versus-host disease. Blood 138, 722–737. https://doi.org/10.1182/blood.2020006375.
- Casazza, R.L., and Lazear, H.M. (2019). Why Is IFN-lambda Less Inflammatory? One IRF Decides. Immunity 51, 415–417. https://doi.org/10.1016/j.immuni.2019.08.019.
- Sohn, S.Y., Hearing, J., Mugavero, J., Kirillov, V., Gorbunova, E., Helminiak, L., Mishra, S., Mackow, E., Hearing, P., Reich, N.C., and Kim, H.K. (2021). Interferon-Lambda Intranasal Protection and Differential Sex Pathology in a Murine Model of SARS-CoV-2 Infection. mBio *12*, e0275621. https://doi.org/10.1128/mBio.02756-21.
- Israelow, B., Song, E., Mao, T., Lu, P., Meir, A., Liu, F., Alfajaro, M.M., Wei, J., Dong, H., Homer, R.J., et al. (2020). Mouse model of SARS-CoV-2 reveals inflammatory role of type I interferon signaling. J. Exp. Med. 217, e20201241. https://doi.org/10.1084/jem.20201241.
- Yang, S., Cao, L., Xu, W., Xu, T., Zheng, B., Ji, Y., Huang, S., Liu, L., Du, J., Peng, H., et al. (2022). Comparison of model-specific histopathology in mouse models of COVID-19. J. Med. Virol. 94, 3605–3612. https://doi. org/10.1002/jmv.27747.
- Dijkman, R., Verma, A.K., Selvaraj, M., Ghimire, R., Gad, H.H., Hartmann, R., More, S., Perlman, S., Thiel, V., and Channappanavar, R. (2022). Effective Interferon Lambda Treatment Regimen To Control Lethal MERS-CoV Infection in Mice. J. Virol. 96, e0036422. https://doi.org/10.1128/jvi. 00364-22.
- Galligan, C.L., Murooka, T.T., Rahbar, R., Baig, E., Majchrzak-Kita, B., and Fish, E.N. (2006). Interferons and viruses: signaling for supremacy. Immunol. Res. 35, 27–40. https://doi.org/10.1385/IR:35:1:27.
- Wang, B.X., and Fish, E.N. (2012). The yin and yang of viruses and interferons. Trends Immunol. 33, 190–197. https://doi.org/10.1016/j.it.2012. 01.004.
- Loo, Y.M., and Gale, M., Jr. (2007). Viral regulation and evasion of the host response. Curr. Top. Microbiol. Immunol. 316, 295–313. https://doi.org/ 10.1007/978-3-540-71329-6\_14.
- Jackson, C.B., Farzan, M., Chen, B., and Choe, H. (2022). Mechanisms of SARS-CoV-2 entry into cells. Nat. Rev. Mol. Cell Biol. 23, 3–20. https://doi. org/10.1038/s41580-021-00418-x.
- Pfaender, S., Mar, K.B., Michailidis, E., Kratzel, A., Boys, I.N., V'Kovski, P., Fan, W., Kelly, J.N., Hirt, D., Ebert, N., et al. (2020). LY6E impairs coronavirus fusion and confers immune control of viral disease. Nat. Microbiol. 5, 1330–1339. https://doi.org/10.1038/s41564-020-0769-y.
- Zhao, X., Zheng, S., Chen, D., Zheng, M., Li, X., Li, G., Lin, H., Chang, J., Zeng, H., and Guo, J.T. (2020). LY6E Restricts Entry of Human Coronaviruses, Including Currently Pandemic SARS-CoV-2. J. Virol. 94, e00562-20. https://doi.org/10.1128/JVI.00562-20.
- Yu, J., Liang, C., and Liu, S.L. (2017). Interferon-inducible LY6E Protein Promotes HIV-1 Infection. J. Biol. Chem. 292, 4674–4685. https://doi. org/10.1074/jbc.M116.755819.
- Mar, K.B., Rinkenberger, N.R., Boys, I.N., Eitson, J.L., McDougal, M.B., Richardson, R.B., and Schoggins, J.W. (2018). LY6E mediates an evolutionarily conserved enhancement of virus infection by targeting a late entry step. Nat. Commun. 9, 3603. https://doi.org/10.1038/s41467-018-06000-y.
- Lesage, S., Chazal, M., Beauclair, G., Batalie, D., Cerboni, S., Couderc, E., Lescure, A., Del Nery, E., Tangy, F., Martin, A., et al. (2022). Discovery of

Genes that Modulate Flavivirus Replication in an Interferon-Dependent Manner. J. Mol. Biol. 434, 167277. https://doi.org/10.1016/j.jmb.2021. 167277.

- 45. Bruchez, A., Sha, K., Johnson, J., Chen, L., Stefani, C., McConnell, H., Gaucherand, L., Prins, R., Matreyek, K.A., Hume, A.J., et al. (2020). MHC class II transactivator CIITA induces cell resistance to Ebola virus and SARS-like coronaviruses. Science 370, 241–247. https://doi.org/10. 1126/science.abb3753.
- Kenney, A.D., Zani, A., Kawahara, J., Eddy, A.C., Wang, X.L., Mahesh, K.C., Lu, M., Thomas, J., Kohlmeier, J.E., Suthar, M.S., et al. (2023). Interferon-induced transmembrane protein 3 (IFITM3) limits lethality of SARS-CoV-2 in mice. EMBO Rep. 24, e56660. https://doi.org/10.15252/embr. 202256660.
- Xu, F., Wang, G., Zhao, F., Huang, Y., Fan, Z., Mei, S., Xie, Y., Wei, L., Hu, Y., Wang, C., et al. (2022). IFITM3 Inhibits SARS-CoV-2 Infection and Is Associated with COVID-19 Susceptibility. Viruses 14. https://doi.org/10. 3390/v14112553.
- Jiménez-Munguía, I., Beaven, A.H., Blank, P.S., Sodt, A.J., and Zimmerberg, J. (2022). Interferon-induced transmembrane protein 3 (IFITM3) and its antiviral activity. Curr. Opin. Struct. Biol. 77, 102467. https://doi. org/10.1016/j.sbi.2022.102467.
- Gholami, M., Sakhaee, F., Sotoodehnejadnematalahi, F., Zamani, M.S., Ahmadi, I., Anvari, E., and Fateh, A. (2022). Increased risk of COVID-19 mortality rate in IFITM3 rs6598045 G allele carriers infected by SARS-CoV-2 delta variant. Hum. Genomics *16*, 60. https://doi.org/10.1186/ s40246-022-00434-8.
- Kristiansen, H., Gad, H.H., Eskildsen-Larsen, S., Despres, P., and Hartmann, R. (2011). The Oligoadenylate Synthetase Family: An Ancient Protein Family with Multiple Antiviral Activities. J. Interferon Cytokine Res. 31, 41–47. https://doi.org/10.1089/jir.2010.0107.
- Morin, B., Rabah, N., Boretto-Soler, J., Tolou, H., Alvarez, K., and Canard, B. (2010). High yield synthesis, purification and characterisation of the RNase L activators 5'-triphosphate 2'-5'-oligoadenylates. Antiviral Res. 87, 345–352. https://doi.org/10.1016/j.antiviral.2010.06.003.
- Chakrabarti, A., Jha, B.K., and Silverman, R.H. (2011). New Insights into the Role of RNase L in Innate Immunity. J. Interferon Cytokine Res. *31*, 49–57. https://doi.org/10.1089/jir.2010.0120.
- Drappier, M., and Michiels, T. (2015). Inhibition of the OAS/RNase L pathway by viruses. Curr. Opin. Virol. 15, 19–26. https://doi.org/10. 1016/j.coviro.2015.07.002.
- Schwerk, J., Soveg, F.W., Ryan, A.P., Thomas, K.R., Hatfield, L.D., Ozarkar, S., Forero, A., Kell, A.M., Roby, J.A., So, L., et al. (2019). RNA-binding protein isoforms ZAP-S and ZAP-L have distinct antiviral and immune resolution functions. Nat. Immunol. 20, 1610–1620. https://doi.org/10.1038/ s41590-019-0527-6.
- Soveg, F.W., Schwerk, J., Gokhale, N.S., Cerosaletti, K., Smith, J.R., Pairo-Castineira, E., Kell, A.M., Forero, A., Zaver, S.A., Esser-Nobis, K., et al. (2021). Endomembrane targeting of human OAS1 p46 augments antiviral activity. Elife *10*, e71047. https://doi.org/10.7554/eLife.71047.
- Wickenhagen, A., Sugrue, E., Lytras, S., Kuchi, S., Noerenberg, M., Turnbull, M.L., Loney, C., Herder, V., Allan, J., Jarmson, I., et al. (2021). A prenylated dsRNA sensor protects against severe COVID-19. Science 374, eabj3624. https://doi.org/10.1126/science.abj3624.
- Huffman, J.E., Butler-Laporte, G., Khan, A., Pairo-Castineira, E., Drivas, T.G., Peloso, G.M., Nakanishi, T., COVID-19 Host Genetics Initiative, Ganna, A., Verma, A., et al. (2022). Multi-ancestry fine mapping implicates OAS1 splicing in risk of severe COVID-19. Nat. Genet. 54, 125–127. https://doi.org/10.1038/s41588-021-00996-8.
- Pairo-Castineira, E., Clohisey, S., Klaric, L., Bretherick, A.D., Rawlik, K., Pasko, D., Walker, S., Parkinson, N., Fourman, M.H., Russell, C.D., et al. (2021). Genetic mechanisms of critical illness in COVID-19. Nature 591, 92–98. https://doi.org/10.1038/s41586-020-03065-y.
- Zhang, Q., Pizzorno, A., Miorin, L., Bastard, P., Gervais, A., Le Voyer, T., Bizien, L., Manry, J., Rosain, J., Philippot, Q., et al. (2022). Autoantibodies against type I IFNs in patients with critical influenza pneumonia. J. Exp. Med. 219, e20220514. https://doi.org/10.1084/jem.20220514.

### CellPress

- Bastard, P., Vazquez, S., Liu, J., Laurie, M.T., Wang, C.Y., Gervais, A., Le Voyer, T., Bizien, L., Zamecnik, C., Philippot, Q., et al. (2022). Vaccine breakthrough hypoxemic COVID-19 pneumonia in patients with auto-Abs neutralizing type I IFNs. Sci. Immunol.eabp8966 https://doi.org/10. 1126/sciimmunol.abp8966.
- Manry, J., Bastard, P., Gervais, A., Le Voyer, T., Rosain, J., Philippot, Q., Michailidis, E., Hoffmann, H.H., Eto, S., Garcia-Prat, M., et al. (2022). The risk of COVID-19 death is much greater and age dependent with type I IFN autoantibodies. Proc. Natl. Acad. Sci. USA *119*, e2200413119. https://doi. org/10.1073/pnas.2200413119.
- Nchioua, R., Kmiec, D., Müller, J.A., Conzelmann, C., Groß, R., Swanson, C.M., Neil, S.J.D., Stenger, S., Sauter, D., Münch, J., et al. (2020). SARS-CoV-2 Is Restricted by Zinc Finger Antiviral Protein despite Preadaptation to the Low-CpG Environment in Humans. mBio 11, e01930-20. https://doi. org/10.1128/mBio.01930-20.
- Zimmer, M.M., Kibe, A., Rand, U., Pekarek, L., Ye, L., Buck, S., Smyth, R.P., Cicin-Sain, L., and Caliskan, N. (2021). The short isoform of the host antiviral protein ZAP acts as an inhibitor of SARS-CoV-2 programmed ribosomal frameshifting. Nat. Commun. 12, 7193. https://doi.org/10.1038/ s41467-021-27431-0.
- Song, J., Chow, R.D., Peña-Hernández, M.A., Zhang, L., Loeb, S.A., So, E.Y., Liang, O.D., Ren, P., Chen, S., Wilen, C.B., and Lee, S. (2022). LRRC15 inhibits SARS-CoV-2 cellular entry in trans. PLoS Biol. 20, e3001805. https://doi.org/10.1371/journal.pbio.3001805.
- 65. Kratzel, A., Kelly, J.N., V'Kovski, P., Portmann, J., Brüggemann, Y., Todt, D., Ebert, N., Shrestha, N., Plattet, P., Staab-Weijnitz, C.A., et al. (2021). A genome-wide CRISPR screen identifies interactors of the autophagy pathway as conserved coronavirus targets. PLoS Biol. 19, e3001490. https://doi.org/10.1371/journal.pbio.3001490.
- Zhu, Y., Feng, F., Hu, G., Wang, Y., Yu, Y., Zhu, Y., Xu, W., Cai, X., Sun, Z., Han, W., et al. (2021). A genome-wide CRISPR screen identifies host factors that regulate SARS-CoV-2 entry. Nat. Commun. *12*, 961. https://doi. org/10.1038/s41467-021-21213-4.
- Mac Kain, A., Maarifi, G., Aicher, S.M., Arhel, N., Baidaliuk, A., Munier, S., Donati, F., Vallet, T., Tran, Q.D., Hardy, A., et al. (2022). Identification of DAXX as a restriction factor of SARS-CoV-2 through a CRISPR/Cas9 screen. Nat. Commun. *13*, 2442. https://doi.org/10.1038/s41467-022-30134-9.
- Israeli, M., Finkel, Y., Yahalom-Ronen, Y., Paran, N., Chitlaru, T., Israeli, O., Cohen-Gihon, I., Aftalion, M., Falach, R., Rotem, S., et al. (2022). Genomewide CRISPR screens identify GATA6 as a proviral host factor for SARS-CoV-2 via modulation of ACE2. Nat. Commun. 13, 2237. https://doi.org/ 10.1038/s41467-022-29896-z.
- Ugalde, A.P., Bretones, G., Rodríguez, D., Quesada, V., Llorente, F., Fernández-Delgado, R., Jiménez-Clavero, M.Á., Vázquez, J., Calvo, E., Tamargo-Gómez, I., et al. (2022). Autophagy-linked plasma and lysosomal membrane protein PLAC8 is a key host factor for SARS-CoV-2 entry into human cells. EMBO J. 41, e110727. https://doi.org/10.15252/embj. 2022110727.
- Hoffmann, H.H., Sánchez-Rivera, F.J., Schneider, W.M., Luna, J.M., Soto-Feliciano, Y.M., Ashbrook, A.W., Le Pen, J., Leal, A.A., Ricardo-Lax, I., Michailidis, E., et al. (2021). Functional interrogation of a SARS-CoV-2 host protein interactome identifies unique and shared coronavirus host factors. Cell Host Microbe 29, 267–280.e5. https://doi.org/10.1016/j.chom.2020. 12.009.
- Wei, J., Alfajaro, M.M., DeWeirdt, P.C., Hanna, R.E., Lu-Culligan, W.J., Cai, W.L., Strine, M.S., Zhang, S.M., Graziano, V.R., Schmitz, C.O., et al. (2021). Genome-wide CRISPR Screens Reveal Host Factors Critical for SARS-CoV-2 Infection. Cell 184, 76–91.e13. https://doi.org/10.1016/j. cell.2020.10.028.
- Wang, R., Simoneau, C.R., Kulsuptrakul, J., Bouhaddou, M., Travisano, K.A., Hayashi, J.M., Carlson-Stevermer, J., Zengel, J.R., Richards, C.M., Fozouni, P., et al. (2021). Genetic Screens Identify Host Factors for SARS-CoV-2 and Common Cold Coronaviruses. Cell 184, 106–119.e14. https://doi.org/10.1016/j.cell.2020.12.004.
- Daniloski, Z., Jordan, T.X., Wessels, H.H., Hoagland, D.A., Kasela, S., Legut, M., Maniatis, S., Mimitou, E.P., Lu, L., Geller, E., et al. (2021). Identification of Required Host Factors for SARS-CoV-2 Infection in Human Cells. Cell 184, 92–105.e16. https://doi.org/10.1016/j.cell.2020.10.030.

Perspective

Immunity

- Crouse, J., Kalinke, U., and Oxenius, A. (2015). Regulation of antiviral T cell responses by type I interferons. Nat. Rev. Immunol. 15, 231–242. https:// doi.org/10.1038/nri3806.
- Leist, S.R., Dinnon, K.H., 3rd, Schäfer, A., Tse, L.V., Okuda, K., Hou, Y.J., West, A., Edwards, C.E., Sanders, W., Fritch, E.J., et al. (2020). A Mouse-Adapted SARS-CoV-2 Induces Acute Lung Injury and Mortality in Standard Laboratory Mice. Cell 183, 1070–1085.e12. https://doi.org/10. 1016/j.cell.2020.09.050.
- 76. Lopez, J., Mommert, M., Mouton, W., Pizzorno, A., Brengel-Pesce, K., Mezidi, M., Villard, M., Lina, B., Richard, J.C., Fassier, J.B., et al. (2021). Early nasal type I IFN immunity against SARS-CoV-2 is compromised in patients with autoantibodies against type I IFNs. J. Exp. Med. 218, e20211211. https://doi.org/10.1084/jem.20211211.
- 77. Bastard, P., Rosen, L.B., Zhang, Q., Michailidis, E., Hoffmann, H.H., Zhang, Y., Dorgham, K., Philippot, Q., Rosain, J., Béziat, V., et al. (2020). Autoantibodies against type I IFNs in patients with life-threatening COVID-19. Science 370, eabd4585. https://doi.org/10.1126/science. abd4585.
- Solanich, X., Rigo-Bonnin, R., Gumucio, V.D., Bastard, P., Rosain, J., Philippot, Q., Perez-Fernandez, X.L., Fuset-Cabanes, M.P., Gordillo-Benitez, M.Á., Suarez-Cuartin, G., et al. (2021). Pre-existing Autoantibodies Neutralizing High Concentrations of Type I Interferons in Almost 10% of COVID-19 Patients Admitted to Intensive Care in Barcelona. J. Clin. Immunol. 41, 1733–1744. https://doi.org/10.1007/s10875-021-01136-x.
- Hao, L., Hsiang, T.Y., Dalmat, R.R., Ireton, R., Morton, J.F., Stokes, C., Netland, J., Hale, M., Thouvenel, C., Wald, A., et al. (2023). Dynamics of SARS-CoV-2 VOC Neutralization and Novel mAb Reveal Protection against Omicron. Viruses 15. https://doi.org/10.3390/v15020530.
- Brown, B., Ojha, V., Fricke, I., Al-Sheboul, S.A., Imarogbe, C., Gravier, T., Green, M., Peterson, L., Koutsaroff, I.P., Demir, A., et al. (2023). Innate and Adaptive Immunity during SARS-CoV-2 Infection: Biomolecular Cellular Markers and Mechanisms. Vaccines 11. https://doi.org/10.3390/ vaccines11020408.
- Spinardi, J.R., and Srivastava, A. (2023). Hybrid Immunity to SARS-CoV-2 from Infection and Vaccination-Evidence Synthesis and Implications for New COVID-19 Vaccines. Biomedicines *11*, 370. https://doi.org/10. 3390/biomedicines11020370.
- Lai, J.Y., Ho, J.X., Kow, A.S.F., Liang, G., Tham, C.L., Ho, Y.C., and Lee, M.T. (2023). Interferon therapy and its association with depressive disorders - A review. Front. Immunol. *14*, 1048592. https://doi.org/10.3389/ fimmu.2023.1048592.
- Chang, L.W., Liranzo, M., and Bergfeld, W.F. (1995). Cutaneous side effects associated with interferon-alpha therapy: a review. Cutis 56, 144.
- Ryoo, S., Koh, D.H., Yu, S.Y., Choi, M., Huh, K., Yeom, J.S., and Heo, J.Y. (2023). Clinical efficacy and safety of interferon (Type I and Type III) therapy in patients with COVID-19: A systematic review and meta-analysis of randomized controlled trials. PLoS One 18, e0272826. https://doi.org/10. 1371/journal.pone.0272826.
- Chen, W.C., Hsu, C.K., Chen, C.Y., Lai, C.C., Hung, S.H., and Lin, W.T. (2022). Clinical efficacy and safety of interferon-beta-containing regimens in the treatment of patients with COVID-19: a systematic review and metaanalysis of randomized controlled trials. Expert Rev. Anti Infect. Ther. 20, 741–747. https://doi.org/10.1080/14787210.2022.2004118.
- Buchynskyi, M., Kamyshna, I., Lyubomirskaya, K., Moshynets, O., Kobyliak, N., Oksenych, V., and Kamyshnyi, A. (2023). Efficacy of interferon alpha for the treatment of hospitalized patients with COVID-19: A metaanalysis. Front. Immunol. *14*, 1069894. https://doi.org/10.3389/fimmu. 2023.1069894.
- Feld, J.J., Kandel, C., Biondi, M.J., Kozak, R.A., Zahoor, M.A., Lemieux, C., Borgia, S.M., Boggild, A.K., Powis, J., McCready, J., et al. (2021). Peginterferon lambda for the treatment of outpatients with COVID-19: a phase 2, placebo-controlled randomised trial. Lancet Respir. Med. 9, 498–510. https://doi.org/10.1016/S2213-2600(20)30566-X.
- Reis, G., Moreira Silva, E.A.S., Medeiros Silva, D.C., Thabane, L., Campos, V.H.S., Ferreira, T.S., Santos, C.V.Q., Nogueira, A.M.R., Almeida, A.P.F.G., Savassi, L.C.M., et al. (2023). Early Treatment with Pegylated Interferon Lambda for Covid-19. N. Engl. J. Med. 388, 518–528. https:// doi.org/10.1056/NEJMoa2209760.